# MED13L-related intellectual disability: involvement of missense variants and delineation of the phenotype 

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#### Abstract

Molecular anomalies in MED13L, leading to haploinsufficiency, have been reported in patients with moderate to severe intellectual disability (ID) and distinct facial features, with or without congenital heart defects. Phenotype of the patients was referred to "MED13L haploinsufficiency syndrome." Missense variants in MED13L were already previously described to cause the MED13L-related syndrome, but only in a limited number of patients. Here we report 36 patients with MED13L molecular anomaly, recruited through an international collaboration between centers of expertise for developmental anomalies. All patients presented with intellectual disability and severe language impairment. Hypotonia, ataxia, and recognizable facial gestalt were frequent findings, but not congenital heart defects. We identified seven de novo missense variations, in addition to proteintruncating variants and intragenic deletions. Missense variants clustered in two mutation hot-spots, i.e., exons 15-17 and 25-31. We found that patients carrying missense mutations had more frequently epilepsy and showed a more severe phenotype. This study ascertains missense variations in $M E D 13 L$ as a cause for $M E D 13 L$-related intellectual disability and improves the clinical delineation of the condition.


Keywords MED13L • Intellectual disability • Mediator complex • Cardiopathy

## Introduction

Mediator is a large coregulator complex conserved from yeast to humans. The complex has emerged as a master coordinator of cell lineage determination, integrating signaling from various transcription factors, epigenetic regulators and non-coding RNAs [1]. In response to various

[^0]stimuli, mediator undergoes conformational changes and creates a DNA loop between activated enhancer elements and promoter, notably through interactions with cohesins [1]. Mediator physically bridges transcription factors bound at enhancer elements with the RNA polymerase II transcription machinery at core promoter regions [1]. Mediator is organized into four modules, i.e., the tail-, the middle-, the head-, and the CDK8-kinase module [2]. In vertebrates, the latter module is composed of CCNT1 and three additional proteins: CDK8, MED12, and MED13; or their respective paralogs: CDK19, MED12L, and MED13L [3]. Disease-causing variations have been identified in genes encoding the CDK8module proteins. MED12 variants cause syndromic intellectual disabilities (ID), namely Opitz-Kaveggia syndrome
(MIM \#305450), Lujan-Fryns syndrome (MIM \#309520), and Ohdo syndrome (MIM \#300895) [4]. CDK19 interruption by a translocation breakpoint has been found to cause moderate ID with microcephaly and retinal folds in a patient [5]. Recently, MED13L haploinsufficiency have been identified in patients with moderate to severe ID, hypotonia, and distinctive facial gestalt (OMIM \#616789) [6-10]. The recognizable syndrome was delineated by Asadollahi et al. [7] and broadened by further reports $[6,8,10-15]$. The gene is located on chromosome 12 q 24.21 and encodes MED13L (alias TRAP240L), expressed in heart and brain tissues [9]. Originally, the interruption of $M E D 13 L$ by a translocation breakpoint was identified in a patient with dextro-loop transposition of the great arteries (dTGA- MIM \#608808) and intellectual disability (ID). Given that association, a cohort of 97 individuals with isolated dTGA was screened for $M E D 13 L$ sequence variations. Rare heterozygous missense variants were identified in four patients [9]. Familial segregation was not available for three variants and showed that the remaining variant was inherited from a healthy parent [9]. Updated annotations of the four variations showed that the variant c.2056A>C was reported 472 times in GnomAD database (http://gnomad. broadinstitute.org/). Variants c.752A $>\mathrm{G}$ and $\mathrm{c} .6068 \mathrm{~A}>\mathrm{G}$ were reported in GnomAD database once and the variant c. $5615 \mathrm{G}>\mathrm{A}$ was reported once in 1000G database (http:// www.internationalgenome.org/). Therefore, clinical relevance of these variants remains unclear. It was hypothesized that missense variants were associated with congenital heart defects (CHDs), particularly dTGA, without intellectual disability [10].

To date, 33 additional patients with a $M E D 13 L$ variants or intragenic deletion were reported $[6-8,10,12,13$, 15-22]. The DDD studies identified at least 19 patients with a MED13L variant, highlighting MED13L as one of the most common ID-causing gene [14, 23]. Variants were either identified by targeted sequencing, indicating that the condition could be suspected prior to the molecular analysis, or by exome sequencing [12]. Strikingly, no further dTGA was found and all patients presented with ID, characteristic facial gestalt, and less commonly aspecific CHD in 6/25 cases (patent foramen ovale, Fallot tetralogy, pulmonary atresia). To our knowledge, seven missense variants were identified in 11 patients, but the lack of precise clinical data in most of them precluded clarification of their clinical relevance or possible genotypephenotype correlation [8, 13, 14, 17]. Here, we report on 36 patients with $M E D 13 L$ variations affecting its function, including seven missense variants in nine patients. We aim to better delineate the phenotype and discuss possible genotype-phenotype correlation.

## Subjects and methods

## Patients

Thirty-six patients from 35 families were recruited through an international collaboration between centers of expertise for developmental anomalies. All patients were clinically examined by a clinical geneticist. Two patients have been published previously, but more detailed information was reported here: P7 (ref. [19]) and P8 (ref. [18]). Informed consents were obtained for genetic tests, data sharing, and publication of patients' photographs.

## Genetic analyses

Molecular investigations were performed in different diagnostic laboratories according to their routine procedure regarding testing in patients affected with ID. MED $13 L$ intragenic deletions were identified by array-CGH. MED $13 L$ sequence variants were identified by either next generation sequencing of custom gene panels designed for ID (P5, P6, P7, P8, P9, P10, P11, P23, P25, P26, P27, and P31) [19, 24] or by whole exome sequencing for the other patients. Missense variations were evaluated using the Alamut interface (Interactive Biosoftware, Rouen, France). Pathogenicity scores were predicted in silico with SIFT (http://sift.jcvi.org), PolyPhen2 (http://genetics.bwh.harvard.edu/pph2/), and MutationTaster (http://www.mutationtaster.org/) softwares. All coordinates are provided for NM_015335.4 transcript in hg19 (genome build: GRCh37) and NP_056150.1 protein. Variant data have been submitted to ClinVar (https://www.ncbi.nlm.nih. gov/clinvar/).

## Results

## MED13L molecular anomalies

MED13L intragenic deletions, ranging in size from 47 to 200 kb , were identified in five patients (P1, P2, P3, P4, and P24) (Fig. 1 and Table 1). When available, parental segregation showed that these occurred de novo. We identified a $M E D 13 L$ sequence variants in 31 patients from 30 families. In one family, recurrence in two sibs (P12 and P13) was observed, presumptively due to parental germinal mosaicism. Protein-truncating variants were identified in 27 patients and were distributed all over the gene (Fig. 1a). One proteintruncating variant, c.1708_1709del, was identified in two unrelated patients (P7-P15). Four variants were predicted to affect splicing (c. $5588+1 \mathrm{G}>\mathrm{A}-\mathrm{c} .1009+1 \mathrm{G}>\mathrm{C}$ and $\mathrm{c} .2345-$ $3 \mathrm{C}>\mathrm{G}$ and $\mathrm{c} .6225+1 \mathrm{G}>\mathrm{A})$. Variant $\mathrm{c} .1009+1 \mathrm{G}>\mathrm{C}$ was identified in two unrelated patients (P25-P30). Seven likely


Fig. 1 a Summary of $M E D 13 L$ variants reported in the literature and in our cohort. Truncating variants are represented above the gene and missense variants under the gene. Asterisks indicate recurrent missense variants. Inter-species conservation of MED $13 L$ missense variations identified in the literature and in our series is shown. H.s, Homo
pathogenic heterozygous missense variants were identified in nine patients (P14, P20, P21, P22, P23 P28, P32, P33, P35): c. $2597 \mathrm{C}>$ T p.(Pro866Leu); c. $2605 \mathrm{C}>$ T p.(Pro869Ser); c. $2930 \mathrm{C}>$ T p. $(\mathrm{Ala} 977 \mathrm{Val}) ;$ c. $6005 \mathrm{C}>$ T p. $(S e r 2002 \mathrm{Leu})$ c. $6485 \mathrm{C}>$ T p. (Thr2162Met); c.6488C>T p.(Ser2163Leu) c. $6530 \mathrm{C}>$ A p.(Ser2177Tyr) and were absent from GnomAD and ExAC database in well-covered regions. Both missense variants c. $2605 \mathrm{C}>\mathrm{T}$ p. (Pro869Ser) and c. $6488 \mathrm{C}>\mathrm{T}$ p.(Ser2163Leu) were identified in two unrelated patients, respectively in P28-P35 and P21-P23. Patient P20-P14 and P33 carried respectively the previously reported variants
sapiens; M.m, Mus musculus; D.r, Danio rerio. b Schematic representation of $M E D 13 L$ and location of intragenic deletions reported in our cohort (P1-P2-P3-P4 and P24) and in the literature, indicated by horizontal bars
c. $2597 \mathrm{C}>$ T p.(Pro866Leu), c.6005C $>$ T p.(Ser2002Leu), and c.6485C $>$ T p. $(\operatorname{Tr} 2162 \mathrm{Met})[14]$. All missense variations were predicted to be "probably damaging" for PolyPhen2 (score > 0.98 ) and deleterious" for SIFT (score $<0.03$ ). The seven missense variants were predicted to induce substitutions involving Pro866, Pro869, Ala977, Ser2002, Thr2162, Ser2163, and Ser2177 residues, which are highly conserved across vertebrates (PhyloP score 5.69 to 6.18 -Fig. 1a). Except for Ala977, which presented a PhyloP score of 6.18 , residues involved in these substitutions are also conserved in MED13, the MED13L paralog (Fig. 1).
Table 1 Clinical and molecular data of the 36 patients harboring likely pathogenic MED13L variants and comparison with the literature

Table 1 (continued)

| Patient | 11 | 12 | 13 | 14 | 15 | 16 | 17 | 18 | 19 | 20 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Bulbous nasal tip | + | + | + | - | + | - | + | + | + | + |
| Cupid-bow upper lip | + | - | - | - | NA | - | - | NA | + | - |
| Hypotonic open-mouth | + | + | + | + | NA | - | + | + | - | + |
| Thin vermillon border | - | - | - | + | + | + | - | - | - | - |
| Deep philtrum | + | - | - | - | NA | + | - | NA | + | - |
| CHD | - | - | - | - | NA | - | - | - | - | - |
| Miscellaneous | CBH syndrome | Obsesity, cryptorchidism, pyramidal syndrome | Obsesity, pyramidal syndrome | Kidney cysts | Posterior cleft palate | Strabismus, cryptorchidism | Hypopallesthesia | Hirsutism, strabismus | Robin sequence, 3-4-5 Toe clinodactyly, severe scoliosis | Nystagmus, craniosynostosis |
| MED $13 L$ | c.5152_5153del | c. $5588+1 \mathrm{G}>\mathrm{A}$ | c. $5588+1 \mathrm{G}>\mathrm{A}$ | c. $6485 \mathrm{C}>\mathrm{T}$ | c.1708_1709del | c.6284dup | c. $830 \_845 \mathrm{del}$ | c. $2065 \mathrm{C}>\mathrm{T}$ | c.3942_3943del | c. $2597 \mathrm{C}>\mathrm{T}$ |
| molecular anomalies | p.(Met1718Glufs*21) | p.(?) | p.(?) | p.(Thr2162Met) | p.(Ser570Phefs*27) | p.(Ala2096Glyfs*12) | p.(Arg277Gln*5) | p.(Gln689*) | p.(Ile1315Glnfs*49) | p.(Pro866Leu) |
| Parental segregation | de novo | de novo | de novo | de novo | de novo | de novo | de novo | NA | de novo | de novo |


| Patient | 21 | 22 | 23 | 24 | 25 | 26 | 27 | 28 | 29 | 30 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Gender | F | F | F | F | F | M | M | F | M | F |
| Age at first examination (years) | 9 | 8 | 12 | 1.6 | 0.7 | 2.5 | 2.7 | 12 | 2 | 39 |
| DD/ID | + (severe) | + | + (severe) | + | + | + | + | + | + | + |
| Speech delay | + | + | + | + | NA | + | + | + | + | + |
| Speech features | abs. | few words | few words | few words | NA | few words | NA | abs. | few words | few sentences |
| Motor delay | + | + | + | + | + | + | + | + | + | + |
| Walk (months) | 30 | 25 | 26 | Not acquired yet | NA | 23 | abs. | abs. | NA | 20 |
| Hypotonia | + | - | + | + | + | + | + | - | + | - |
| Ataxia | - | - | + | - | NA | + (dysarthria) | NA | - | - | + |
| Seizures | + | - | - | - | - | - | - | + | - | - |
| Autistic features | + | + | - | - | NA | - | - | NA | - | - |
| Behavioral troubles | - | + | - | - | - | + | + | NA | - | + |
| MRI | Normal | Focal cortical dysplasia | NA | NA | NA | Normal | NA | CCH- Hypomyelination | NA | Normal |
| Upslanting palpebral fissures | - | - | + | - | - | + | - | + | + | + |
| Bulbous nasal tip | + | - | + | + | - | - | - | + | + | + |
| Cupid-bow upper lip | - | - | + | + | + | + | - | + | + | + |
| Hypotonic open-mouth | + | - | + | + | + | + | - | + | + | + |
| Thin vermillon border | - | - | + | - | + | + | + | + | + | + |
| Deep philtrum | - | - | + | - | + | + | - | + | + | + |
| CHD | - | - | - | - | Pulmonary vavular stenosis | - | - | - | - | - |
| Miscellaneous |  |  | Vertebral artery occlusion | Bilateral talipes, umbilical hernia |  |  | Bilateral talipes | IUGR clynodactyly- double ureter-CoAo | 5 Toe clinodactyly |  |
| MED 13L | c. $6488 \mathrm{C}>\mathrm{T}$ | c. $2930 \mathrm{C}>\mathrm{T}$ | c. $6488 \mathrm{C}>\mathrm{T}$ | exon 3 to 4 deletion | c. $1009+1 \mathrm{G}>\mathrm{C}$ | c. $2340 \_2343 \mathrm{del}$ |  | c. $2605 \mathrm{C}>\mathrm{T}$ | c. 1903dupA | c. $1009+1 \mathrm{G}>\mathrm{C}$ |

Table 1 (continued)

abs., absent speech; $C C$, CoAo, coarctation of the aorta; Corpus Callosum; $C B H$, Claude Bernard-Horner; $C H D$, congenital heart disease; $D D$, developmental delay; $F$, female; $I D$, intellectual disability; $M$, male; MRI, magnetic resonance imaging; NA, non available; VM, VentriculoMegaly; WM, white matter; IUGR, intrauterin growth retardation


Fig. 2 Morphological features of a selection of patients. a Patients with protein-truncating variants. Core facial features comprises depressed nasal bridge, horizontal eyebrows, full cheeks, and large open mouth. Majority of patients show also cupid-bow upper lip, thin vermilion border, and deep philtrum. b Patients P20-P28 and P35 (at different
age) show atypical facial gestalt with long down-slanting palpebral fissures and everted lower eyelids. c Some patients, notably in infancy have broad, stubby, and tapering fingers. Feet showed long halluces and sandal-gap deformity in some patients. d Photo enlargement of the palpebral features of patients P20-P28 and P35

## Phenotypic findings

## Patient with protein-truncating mutation

Protein-truncating variants were identified in 27 patients. No remarkable prenatal history was reported and birth parameters were normal for all individuals. Motor skills were delayed, median age for independent walking being 25 months (range from 17 to 41 months). One patient did not achieve walking but he was only 32 -month-old. Speech was also severely impaired in most individuals, composed of few words (16/21$52 \%$ ) or even absent ( $5 / 21-24 \%$ ). All patients showed moderate to severe ID. Global hypotonia was observed in 20/25 ( $80 \%$ ) patients. Ataxia was noticed in $9 / 25$ patients ( $36 \%$ ), consisting mainly in dynamic ataxia and dysarthria in four patients. We did not retrieve age at onset of the cerebellar signs, but mean age of the patients with ataxia was 12 years (ranging from 2 to 39 years). One patient presented with
seizures ( $1 / 26-4 \%$ ). Autistic features were noticed in $5 / 21$ ( $24 \%$ ) cases and behavioral troubles in 10/26 (39\%), consisting in aggressive behavior when specified. Brain magnetic resonance (MRI) imaging showed various non-specific anomalies comprising ventriculomegaly, myelination defect, corpus callosum anomaly, or focal cortical dysplasia (Table 1). Majority of the patients shared common facial features with wide open mouth, protruding tongue (without macroglossia), full cheeks, bulbous nasal tip, and horizontal eyebrows. Some patients showed thin vermilion, deep philtrum, and cupid-bow upper lip (Fig. 2 and Table 1). Echocardiography revealed patent foramen ovale in two patients (P1) and pulmonary valvular stenosis in one patient (P25) (Supplemental Table 1).

## Patient with missense variation

Missense variants were identified in nine patients. Intra-Uterine Growth Retardation (IUGR) was observed only in P33. Median
age for independent walking was also 25 months (range from 18 to 30 months), but $4 / 9$ patients ( $44 \%$ ) were not able to walk at the age of examination ( $\mathrm{P} 20-\mathrm{P} 28-\mathrm{P} 33-\mathrm{P} 35$ ). The latter patients either did not achieve independent walking (P28-P33) or achieved independent walking and then lost ability to walk because of worsening of epilepsy (P20-P35). Speech was absent in $5 / 9$ patients ( $56 \%$ ) and composed of few words in $4 / 9$ patients ( $44 \%$ ). Global hypotonia was observed in 6/9 patients ( $67 \%$ ). Ataxia was noticed in $3 / 7$ patients ( $43 \%$ ). Six patients presented with seizures $6 / 9$ patients ( $67 \%$ ), consisting in febrile seizure (P32), late onset infantile spasms (P20) and Lennox-Gastaut syndrome (P35). Abnormal brain magnetic resonance imaging (MRI) showed various non-specific anomalies in $4 / 7$ patients ( $57 \%$ ), comparable with these found in patients with proteintruncating variations (supplemental Table 1). We observed atypical facial features in three patients (P20-P28-P35). They showed long down-slanting palpebral fissures, with everted lower eyelids (Fig. 2d). Echocardiography revealed a patent foramen ovale in one patient (P33) (Table 1).

## Discussion

Here we report on a cohort of 36 patients carrying MED13L anomalies, including two previously published cases, allowing a better delineation of the associated phenotype. All individuals had motor delay, speech delay, and moderate to severe ID. In most patients, language was limited to few words or was even absent. Patients showed various degrees of cerebellar dysfunction. Ataxia was observed in 11/32 patients, and was reported especially in the older cases (P9-P10-P13-P15-P17-P20-P23-P32-P32-P36). Since this feature was under-reported in younger patients, we hypothesize that cerebellar involvement probably worsened with age. No patient showed cerebellar anomaly on brain MRI, but repeated imaging may be needed to explore possible progressive atrophy. MRI identified aspecific features comprising myelination defects, corpus callosum abnormalities, white matter anomalies, ventriculomegaly and focal cortical dysplasia. This study confirms that most patients show a recognizable facial gestalt, which could phenotypically overlap with deletion 1p36 microdeletion syndrome (OMIM\# 607872) in some patients [12]. Core facial features comprise depressed nasal bridge, horizontal eyebrows, full cheeks, and large open mouth [7, 12]. More subtle features like cupid-bow upper lip, thin vermilion border, and deep philtrum can be observed. However, in a few patients, these core facial features were absent (P22-P27). We also noticed that non-recurrent facial features can be associated, especially in patients with a missense variant located in the exon 15 (P20-P28-P35) (Fig. 2b). In our cohort, patients were not suspected with the condition prior to genetic testing, but secondarily facial comparison allowed the description of common features. These data highlight the valuable role of clinical geneticists in the precision of the
phenotype, a critical step to determine the pathogenicity of MED13L-variants.

We found only three patients with CHD, consisting in patent foramen ovale or pulmonary valvular stenosis but no complex CHD (Table 1). Patients carried respectively MED $13 L$ intragenic deletion, splice-site variant, and missense variant (P1-P25-P33), confirming that frequency of complex CHD is less than initially expected and is not correlated with MED13L-missense variants. Concerning patients reported by Muncke et al., showing dTGA and probably no developmental delay [9], it is unlikely that their variants affect the protein function. MED13L is located next to TBX3 and TBX5 genes. These genes respectively cause ulnar-mammary syndrome (OMIM\#181450) and Holt-Oram syndrome (OMIM\#142900), both conditions comprising CHD. Heart-specific enhancers have been identified within regulatory domains of both genes; however, they do not overlap with regions contacting MED13L [25]. Therefore, it is unlikely that $M E D 13 L$ variants could affect cis-regulatory elements controlling $T B X 3$ or $T B X 5$ expression during heart development. Involvement of MED13L in the dTGA of these patients remains to be explained.

We observed clinical variability, notably in patients who carried recurrent variations (P12-P13; P21-P23; P7-P15; P25P30; P28-P35) (Table 1). As suggested by Asadollahi et al., we found that patients with missense variants were more prompt to develop epilepsy, compared to patients with protein-truncating variants ( $4 / 9$ versus $1 / 26$ ) [8]. Severe neurodevelopmental phenotype (absent speech in $5 / 9$ versus $5 / 21$ - non ambulatory $4 / 9$ versus $1 / 21$ ) and malformations are also more frequent (Supplemental Table 1). More precisely, patients P20 and P35 lost the ability to walk consecutively to worsening of epilepsy. They needed gastrostomy tube feeding and P35 had hearing impairment and severe myopia. P33 showed a severe phenotype associating IUGR, absent speech, microcephaly, colobomatous microphthalmia, and never achieved sitting. Moreover, P20-P28 and P35 showed atypical facial features with long palpebral fissures and even everted lower eyelid in P20 and P35 (Fig. 2b). Based on these particular palpebral features, P20 was initially diagnosed with Kabuki syndrome (OMIM \#147920). All these data are supported by clinical features from the 11 patients reported in the literature, carrying missense variants [8, 13, 14, 17]. Clinical details of the patients reported by the DDD study were retrieved from Decipher database (https://decipher.sanger.ac.uk/) and corresponded to patients' ID: \#272205, \#260542, \#262717, \#258131, \#268019, \#262545, \#265953, and \#323183. Among them, four patients had epilepsy, and two patients had IUGR and atypical features were also noted (craniosynostosis, microcephaly, major feeding difficulties, limb malformations). Since patients with missense mutation seem to have a more severe phenotype, we could hypothesize that they induce a dominant negative effect, contrarily to protein-truncating variants. We did not consider the patient reported by Mullegama
et al., who suffered from speech delay, ASD and Mediterranean fever. He carried $M E D 13 L, D E A F 1$, and $M E F V$ variants [26]. Inheritance of the $M E D 13 L$ missense variant $\mathrm{c} .5282 \mathrm{C}>\mathrm{T} \mathrm{p}$. (Prol1761Leu) could not be determined, since the patient was adopted. Polyphen 2 and SIFT software predicted in silico that the variant was respectively benign and tolerated. There is no experimental evidence of the deleterious effect on the function of the protein. Thus, there was not enough evidence to consider the variant as the cause of the neurodevelopmental disorder of the patient.

The seven missense variants identified in this study, as well as the MED13L-missense variants previously reported in the literature, cluster in exons 15-17 and 25-31 (Fig. 1) [8, 13, 14, 17]. Both localizations constitute hot-spots of mutations. As expected, majority of missense variations are recurrent [8]. Previously reported variants c. $2597 \mathrm{C}>$ T p.(Pro866Leu), c. $6005 \mathrm{C}>\mathrm{T}$ p.(Ser2002Leu), c.6485C>T p.(Thr2162Met) were identified in three patients [8, 13, 14, 17]. We identified four novel missense variants in six patients: variant c. $2605 \mathrm{C}>\mathrm{T}$ p.(Pro869Ser) was identified in two patients and as well as variant c.6488C>T p.(Ser2163Leu). Variants c.2930C>T p.(Ala977Val) and c.6530C>A p.(Ser2177Tyr) were not recurrent. One of the MED13/MED13L functions is to physically link the CDK8-module to the core Mediator complex, mainly by interacting with MED19 and CDK8 [27]. Dissociation of the CDK8-module components from the core Mediator is mediated by Mediator-bound MED13/MED13L ubiquitylation and degradation [28]. Both subunits can also relay information from temporal/spatial signals or transcription factors to the RNA polymerase II machinery, thus controlling the expression of specific genes, notably genes involved in Wnt, FGF, and Rb/ E2F pathways [8]. Since all the residues involved in the substitutions are located in highly conserved across vertebrate regions and even conserved in MED13, we can assume that these residues are probably implicated in such mechanisms. Further studies are needed to unravel the deleterious mechanism induced by these molecular changes.

## Conclusion

In this cohort of 36 patients with MED13L-related intellectual disability, we confirmed recognizable facial gestalt and intellectual involvement. We highlighted possible arising of progressive cerebellar signs. We did not confirm congenital heart defects as a major feature of the condition. Patients with missense variant are significantly more at risk to develop epilepsy and seemed to have a more severe phenotype, suggesting possible dominant negative effect of the missense variants. We observed clustering of missense variants in specific domains of the protein. Substitution involving the highly conserved across species residues Asp860, Pro866, Pro869, Gly1899, Ser2002, Thr2162, and Ser2163 were identified in at least two patients. Precise
roles of these domains and specific residues remain to be determined to better understand molecular mechanisms underlying MED13L-related intellectual disability.

## Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

## References

1. Allen BL, Taatjes DJ (2015) The mediator complex: a central integrator of transcription. Nat Rev Mol Cell Biol 16(3):155-166
2. Yin J-w, Wang G (2014) The mediator complex: a master coordinator of transcription and cell lineage development. Development 141(5):977-987
3. Daniels DL (2013) Mutual exclusivity of MED12/MED12L, MED13/13L, and CDK8/19 paralogs revealed within the CDKmediator kinase module. J Proteomics Bioinformat 01(S2)
4. Graham JM, Schwartz CE (2013) MED12 related disorders. Am J Med Genet A 161A(11):2734-2740
5. Mukhopadhyay A, Kramer JM, Merkx G, Lugtenberg D, Smeets DF, Oortveld MAW, Blokland EAW, Agrawal J, Schenck A, van Bokhoven H, Huys E, Schoenmakers EF, van Kessel AG, van Nouhuys CE, Cremers FPM (2010) CDK19 is disrupted in a female patient with bilateral congenital retinal folds, microcephaly and mild mental retardation. Hum Genet 128(3):281-291
6. Adegbola A, Musante L, Callewaert B, Maciel P, Hu H, Isidor B, Picker-Minh S, le Caignec C, Delle Chiaie B, Vanakker O, Menten B, Dheedene A, Bockaert N, Roelens F, Decaestecker K, Silva J, Soares G, Lopes F, Najmabadi H, Kahrizi K, Cox GF, Angus SP, Staropoli JF, Fischer U, Suckow V, Bartsch O, Chess A, Ropers HH, Wienker TF, Hübner C, Kaindl AM, Kalscheuer VM (2015) Redefining the MED13L syndrome. Eur J Hum Genet 23(10): 1308-1317
7. Asadollahi R, Oneda B, Sheth F, Azzarello-Burri S, Baldinger R, Joset P, Latal B, Knirsch W, Desai S, Baumer A, Houge G, Andrieux J, Rauch A (2013) Dosage changes of MED13L further delineate its role in congenital heart defects and intellectual disability. Eur J Hum Genet 21(10):1100-1104
8. Asadollahi R et al. Genotype-phenotype evaluation of MED13L defects in the light of a novel truncating and a recurrent missense mutation. Eur J Med Genet 60(9):451-464
9. Muncke N, Jung C, Rüdiger H, Ulmer H, Roeth R, Hubert A, Goldmuntz E, Driscoll D, Goodship J, Schön K, Rappold G (2003) Missense mutations and gene interruption in PROSIT240, a novel TRAP240-like gene, in patients with congenital heart defect (transposition of the great arteries). Circulation 108(23):2843-2850
10. van Haelst MM, Monroe GR, Duran K, van Binsbergen E, Breur JM, Giltay JC, van Haaften G (2015) Further confirmation of the MED13L haploinsufficiency syndrome. Eur J Hum Genet 23(1): 135-138
11. $2017 / 06 / 17 / 09: 38: 34$
12. Cafiero C, Marangi G, Orteschi D, Ali M, Asaro A, Ponzi E, Moncada A, Ricciardi S, Murdolo M, Mancano G, Contaldo I, Leuzzi V, Battaglia D, Mercuri E, Slavotinek AM, Zollino M (2015) Novel de novo heterozygous loss-of-function variants in MED13L and further delineation of the MED13L haploinsufficiency syndrome. Eur J Hum Genet 23(11):1499-1504
13. Caro-Llopis A, Rosello M, Orellana C, Oltra S, Monfort S, Mayo S, Martinez F (2016) De novo mutations in genes of mediator
complex causing syndromic intellectual disability: mediatorpathy or transcriptomopathy? Pediatr Res 80:809-815
14. Deciphering Developmental Disorders, S (2017) Prevalence and architecture of de novo mutations in developmental disorders. Nature 542(7642):433-438
15. Gordon CT, Chopra M, Oufadem M, Alibeu O, Bras M, Boddaert N, Bole-Feysot C, Nitschké P, Abadie V, Lyonnet S, Amiel J (2018) MED13L loss-of-function variants in two patients with syndromic Pierre Robin sequence. Am J Med Genet A 176(1):181-186
16. Iossifov I, Ronemus M, Levy D, Wang Z, Hakker I, Rosenbaum J, Yamrom B, Lee YH, Narzisi G, Leotta A, Kendall J, Grabowska E, Ma B, Marks S, Rodgers L, Stepansky A, Troge J, Andrews P, Bekritsky M, Pradhan K, Ghiban E, Kramer M, Parla J, Demeter R, Fulton LL, Fulton RS, Magrini VJ, Ye K, Darnell JC, Darnell RB, Mardis ER, Wilson RK, Schatz MC, McCombie WR, Wigler $M$ (2012) De novo gene disruptions in children on the autistic spectrum. Neuron 74(2):285-299
17. Gilissen C, Hehir-Kwa JY, Thung DT, van de Vorst M, van Bon BWM, Willemsen MH, Kwint M, Janssen IM, Hoischen A, Schenck A, Leach R, Klein R, Tearle R, Bo T, Pfundt R, Yntema HG, de Vries BBA, Kleefstra T, Brunner HG, Vissers LELM, Veltman JA (2014) Genome sequencing identifies major causes of severe intellectual disability. Nature 511(7509):344-347
18. Hamdan FF, Srour M, Capo-Chichi JM, Daoud H, Nassif C, Patry L, Massicotte C, Ambalavanan A, Spiegelman D, Diallo O, Henrion E, Dionne-Laporte A, Fougerat A, Pshezhetsky AV, Venkateswaran S, Rouleau GA, Michaud JL (2014) De novo mutations in moderate or severe intellectual disability. PLoS Genet 10(10):e1004772
19. Redin C, Gérard B, Lauer J, Herenger Y, Muller J, Quartier A, Masurel-Paulet A, Willems M, Lesca G, el-Chehadeh S, le Gras S, Vicaire S, Philipps M, Dumas M, Geoffroy V, Feger C, Haumesser N, Alembik Y, Barth M, Bonneau D, Colin E, Dollfus H, Doray B, Delrue MA, Drouin-Garraud V, Flori E, Fradin M, Francannet C, Goldenberg A, Lumbroso S, Mathieu-Dramard M, Martin-Coignard D, Lacombe D, Morin G, Polge A, Sukno S, Thauvin-Robinet C, Thevenon J, Doco-Fenzy M, Genevieve D, Sarda P, Edery P, Isidor B, Jost B, Olivier-Faivre L, Mandel JL, Piton A (2014) Efficient strategy for the molecular diagnosis of intellectual disability using targeted high-throughput sequencing. J Med Genet 51(11):724-736
20. Utami KH, Winata CL, Hillmer AM, Aksoy I, Long HT, Liany H, Chew EG, Mathavan S, Tay SK, Korzh V, Sarda P, Davila S, Cacheux V (2014) Impaired development of neural-crest cell-
derived organs and intellectual disability caused by MED13L haploinsufficiency. Hum Mutat 35(11):1311-1320
21. Wang T, Guo H, Xiong B, Stessman HAF, Wu H, Coe BP, Turner TN, Liu Y, Zhao W, Hoekzema K, Vives L, Xia L, Tang M, Ou J, Chen B, Shen Y, Xun G, Long M, Lin J, Kronenberg ZN, Peng Y, Bai T, Li H, Ke X, Hu Z, Zhao J, Zou X, Xia K, Eichler EE (2016) De novo genic mutations among a Chinese autism spectrum disorder cohort. Nat Commun 7:13316
22. Popp B, Ekici AB, Thiel CT, Hoyer J, Wiesener A, Kraus C, Reis A, Zweier C (2017) Exome Pool-Seq in neurodevelopmental disorders. Eur J Hum Genet 25(12):1364-1376
23. Deciphering Developmental Disorders, S (2015) Large-scale discovery of novel genetic causes of developmental disorders. Nature 519(7542):223-228
24. Lehalle D, Mosca-Boidron AL, Begtrup A, Boute-Benejean O, Charles P, Cho MT, Clarkson A, Devinsky O, Duffourd Y, Duplomb-Jego L, Gérard B, Jacquette A, Kuentz P, MasurelPaulet A, McDougall C, Moutton S, Olivié H, Park SM, Rauch A, Revencu N, Rivière JB, Rubin K, Simonic I, Shears DJ, Smol T, Taylor Tavares AL, Terhal P, Thevenon J, van Gassen K, Vincent-Delorme C, Willemsen MH, Wilson GN, Zackai E, Zweier C, Callier P, Thauvin-Robinet C, Faivre L (2017) STAG1 mutations cause a novel cohesinopathy characterised by unspecific syndromic intellectual disability. J Med Genet 54:479-488
25. van Weerd JH, Badi I, van den Boogaard M, Stefanovic S, van de Werken HJG, Gomez-Velazquez M, Badia-Careaga C, Manzanares M, de Laat W, Barnett P, Christoffels VM (2014) A large permissive regulatory domain exclusively controls Tbx 3 expression in the cardiac conduction system. Circ Res 115(4):432-441
26. Mullegama SV, Jensik P, Li C, Dorrani N, UCLA Clinical Genomics Center, Kantarci S, Blumberg B, Grody WW, Strom SP (2017) Coupling clinical exome sequencing with functional characterization studies to diagnose a patient with familial Mediterranean fever and MED13L haploinsufficiency syndromes. Clin Case Rep 5(6):833-840
27. Tsai KL, Sato S, Tomomori-Sato C, Conaway RC, Conaway JW, Asturias FJ (2013) A conserved mediator-CDK8 kinase module association regulates mediator-RNA polymerase II interaction. Nat Struct Mol Biol 20(5):611-619
28. Davis MA, Larimore EA, Fissel BM, Swanger J, Taatjes DJ, Clurman BE (2013) The SCF-Fbw7 ubiquitin ligase degrades MED13 and MED13L and regulates CDK8 module association with mediator. Genes Dev 27(2):151-156

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